# Quad-Modal Fluorescent Nanothermometers Based On Core/Shell CulnS<sub>2</sub>/ZnS Quantum Dots

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Temperature detection is one of the key issues in intracellular research. Monitoring changes in the temperature allows us to track processes taking place in the cell. Therefore, it is important to find a non-invasive, non-toxic material that will enable temperature detection in individual parts of the cell. Colloidal CIS/ZnS QDs are the focus of interest due to their size-dependent luminescence, broadband absorption, low excitation energy, and lack of heavy metal ions. These properties enable the application of QDs in biological research as nanothermometers and make them a competitive material compared to other systems, e.g., fluorescent polymers, proteins, and upconverting nanoparticles. In our work, we aim to improve the sensitivity of the thermometer, to enable the most accurate temperature measurements.

#### Conclusions

- Our thermometer's readout modes are based on (i) PL intensity, (ii) PL lifetime, (iii) PL peak shift, and (iv) a PL intensity ratio under two specific excitation wavelengths which allowed us to obtain quad-modal fluorescent nanothermometer.
- All modes of our nanothermometer can be readily employed using confocal microscopes.
- ► The highest obtained sensitivity for our nanothermometer is based on the PL intensity mode and is equal to 3.2% this value is the highest sensitivity among inorganic semiconductor QDs.
- ► The new MLR analysis method allowed us to obtain multiparametric temperature readout, which increased our thermometer's sensitivity up to 34%.
- Negligible changes in CIS/ZnS-30 absorbance, indicate that QDs are stable in an aqueous environment for 56 days.
- Temperature-induced changes in PL intensity and PL peak position are reproducible and reversible within 20-60°C temperature range.
- Confocal fluorescent microscope images of HeLa cells incubated with CIS QDs revealed that QDs successfully entered the HeLa cells.



Fig. 1: A: Absorption and second derivative, and B: PL spectra of CIS/ZnS QDs after different ZnS synthesis times. C: Quantum yield of CIS/ZnS QDs with different ZnS synthesis times. The CIS and CIS/ZnS QDs were synthesized as described in Refs. [1,2].

Fig. 2: A: PL spectra excited at 450 nm measured as a function of temperature for CIS/ZnS-30 QDs encapsulated in micelles. B: Temperature dependence of the normalized PL intensity (points). C: PL peak position as a function of temperature (points). D: Temperature dependence of normalized PL decays excited at 400 nm. E: Temperature dependence of PL lifetime ( $\tau_{0.1}$ ).  $\tau_{0.1}$  is defined as the decay time at which the intensity drops by a factor of 10. F: PL spectra after 450 (blue line) and 580 nm (red line) excitation. The blue area denotes the spectrum integration range. G: Normalized integrated PL intensities as a function of temperature, for excitation at 450 nm ( $I_{450}$ , blue circles) and 580 nm ( $I_{580}$ , red triangles). H: Temperature dependence of  $I_{450}/I_{580}$  ratio (blue points). Red lines represent linear fits to the data.

dependent PL processes. Solid upward and downward arrows denote, respectively, excitation and non-radiative recombination. The wavy arrow denotes radiative recombination. Doubleheaded arrows show the barrier heights for hot carrier trapping.

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S/CdS/CdSe

CIS/ZnS-30

## Multiparametric temperature sensing

#### Nanothermometer's sensitivity

We evaluated multiparametric sensitivity for our nanothermometers based on Ref. [4]. In the Multiple Linear Regression (MLR) method, the thermometric parameters (Q<sub>i</sub>) have to vary linearly, so the temperature (T) can be expressed as  $T = \beta_0 + \beta_1 Q_1 + ... + \beta_4 Q_4$ , where the  $\beta_0$  is the intercept and  $\beta_i$  is the slope of each Q<sub>i</sub>. Then, the multiparametric sensitivity can be defined as:  $S_{MLR} = \sqrt{\sum_{i=1}^{4} (Q_i \beta_i)^{-2}}$ .

To compare the nanothermometer's performance, for each sample and readout mode the relative sensitivity was calculated based on the equation:  $S_{\xi} = \left| \frac{1}{Q_{\xi}} \frac{dQ_{\xi}}{dT} \right|$ , where  $\xi$  is *I* as intensity, *E* as PL maximum,  $\tau$  as PL lifetime, or *R* as ratiometric mode.



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(eV)

C: PL intensity and D: PL peak position over 3

heating/cooling cycles (temp. range  $20-60^{\circ}$ C).

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1 2 3

Cycle (number)

Fig. 4: A, C, E, G, I: Readout temperatures vs. the experimental temperatures. B, D, F, H, J: Temperature readout residuals indicating the precision of each mode (given in the legends). K: Sensitivities of the four readout modes and the multiparametric readout based on MLR.





- ► PL spectra of CIS/ZnS QDs encapsulated in micelles are slightly redshifted
- ► Red-shift occurs as a result of changing the CIS/ZnS QDs environment to more polar.
- Fig. 5: A: Scheme of the micelles encapsulation. B: PL spectra for CIS/ZnS QDs dispersed in toluene (solid lines) and CIS/ZnS QDs encapsulated in micelles dispersed in water (dashed lines) with ZnS synthesis times. CIS/ZnS QDs encapsulated in micelles were prepared according to Ref. [3].



## References

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Fig. 8: The confocal microscope images of A-D: HeLa cells and E-G: HeLa cells incubated with  $25\mu g$  of CIS/ZnS-120 QDs. A, E: Overlay image of blue, green, and red channels. B: and F: Images of cells with nuclei stained with Hoechst 33342 dye (blue channel, emission) range 425-475 nm). C: and G: Images of cells with lysosomes stained with Alexa Fluor 488conjugated protein (green channel, emission range 494-525 nm). D: and H: CIS/ZnS-120 QDs (red channel, emission range 590-740 nm). All scale bars correspond to 50  $\mu$ m.

# Acknowlegments

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